RESEARCH PAPER

Pharmacological characterization of the new histamine H₄ receptor agonist VUF 8430

Herman D Lim¹, Maristella Adami², Elena Guaita², Thomas Werfel³, Rogier A Smits¹, Iwan JP de Esch¹, Remko A Bakker^{1*}, Ralf Gutzmer³, Gabriella Coruzzi² and Rob Leurs¹

¹Leiden/Amsterdam Center for Drug Research, Division of Medicinal Chemistry, Faculty of Science, Vrije Universiteit Amsterdam, Amsterdam, The Netherlands, ²Department of Human Anatomy, Pharmacology and Forensic Medicine, Section of Pharmacology, University of Parma, Parma, Italy, and ³Department of Dermatology and Allergy Research, Hannover Medical University, Hannover, Germany

Background and purpose: We compare the pharmacological profiles of a new histamine H₄ receptor agonist 2-(2-guanidinoethyl)isothiourea (VUF 8430) with that of a previously described H₄ receptor agonist, 4-methylhistamine.

Experimental approach: Radioligand binding and functional assays were performed using histamine H_4 receptors expressed in mammalian cell lines. Compounds were also evaluated *ex vivo* in monocyte-derived dendritic cells endogenously expressing H_4 receptors and *in vivo* in anaesthetized rats for gastric acid secretion activity.

Key results: Both VUF 8430 and 4-methylhistamine were full agonists at human H₄ receptors with lower affinity at rat and mouse H₄ receptors. Both compounds induced chemotaxis of monocyte-derived dendritic cells. VUF 8430 also showed reasonable affinity and was a full agonist at the H₃ receptor. Agmatine is a metabolite of arginine, structurally related to VUF 8430, and was a H₄ receptor agonist with micromolar affinity. At histamine H₃ receptors, agmatine was a full agonist, whereas 4-methylhistamine was an agonist only at high concentrations. Both VUF 8430 and agmatine were inactive at H₁ and H₂ receptors, whereas 4-methylhistamine is as active as histamine at H₂ receptors. *In vivo*, VUF 8430 only caused a weak secretion of gastric acid mediated by H₂ receptors, whereas 4-methylhistamine, dimaprit, histamine and amthamine, at equimolar doses, induced 2.5- to 6-fold higher output than VUF 8430.

Conclusions and implications: Our results suggest complementary use of 4-methylhistamine and VUF 8430 as H_4 receptor agonists. Along with H_4 receptor antagonists, both agonists can serve as useful pharmacological tools in studies of histamine H_4 receptors.

British Journal of Pharmacology (2009) 157, 34-43; doi:10.1111/j.1476-5381.2009.00200.x

Keywords: histamine H₄ receptor; agonist; 4-methylhistamine; VUF 8430; agmatine; chemotaxis; gastric acid secretion

Abbreviations: CRE, cAMP responsive element; JNJ 7777120, 1-[(5-chloro-1*H*-indol-2-yl)carbonyl]-4-methylpiperazine; MoDC, monocyte-derived dendritic cell; VUF 8430, 2-(2-quanidinoethyl)isothiourea

Introduction

Histamine is a chemical mediator that controls many physiological functions through the interaction with four histamine receptor subtypes, which are all members of the large multigene family of G-protein coupled receptors (Parsons and

Ganellin, 2006). The histamine H₁ and H₂ receptors represent very successful therapeutic targets (Parsons and Ganellin, 2006; receptor nomenclature follows Alexander *et al.*, 2008), while the H₃ receptor and H₄ receptor have emerged as potential targets for future treatment of central nervous system (CNS) disorders (Leurs *et al.*, 2005) and inflammatory diseases (de Esch *et al.*, 2005; Lim *et al.*, 2006b) respectively. The potential therapeutic use is strongly related to the relative selective tissue distribution of both the H₃ receptor and H₄ receptor. Whereas the H₃ receptor is mainly present in the nervous system, the H₄ receptor is primarily localized on haematopoietic cells (Oda *et al.*, 2000; Liu *et al.*, 2001a; Morse *et al.*, 2001; Zhu *et al.*, 2001). The H₄ receptor has been demonstrated to be involved in the chemotaxis of mast cells, eosinophils and monocyte-derived dendritic cells (MoDCs)

Correspondence: Rob Leurs, Leiden/Amsterdam Center for Drug Research, Division of Medicinal Chemistry, Faculty of Science, Vrije Universiteit Amsterdam, De Boelelaan 1083, Amsterdam 1081 HV, The Netherlands. E-mail: r.leurs@few.vu.nl

Received 5 September 2008; revised 14 October 2008; accepted 26 November 2008

^{*}Current address: Boehringer Ingelheim Pharma GmbH & Co.KG, Biberach, Germany.

(Hofstra *et al.*, 2003; Ling *et al.*, 2004; Gutzmer *et al.*, 2005), to control mediator production, such as interleukin (IL)-16 release by human CD8+ T cells (Gantner *et al.*, 2002), leukotriene B₄ production by mast cells (Takeshita *et al.*, 2003; Thurmond *et al.*, 2004), and suppression of IL-12p70 production by MoDCs (Gutzmer *et al.*, 2005). Various studies suggest that the H₄ receptor is a potential new drug target for inflammatory diseases, including chronic allergy, asthma, atopic dermatitis and inflammatory bowel diseases (Thurmond *et al.*, 2004; Dunford *et al.*, 2006; 2007). Moreover, the H₄ receptor has been detected in primary synovial culture obtained from rheumatoid arthritis patients and colorectal cancer tissues, suggesting a possible role for H₄ receptors in these diseases as well (Cianchi *et al.*, 2005; Ohki *et al.*, 2007).

Despite a growing body of evidence, validation of the histamine H₄ receptor as a drug target is mandatory. For this purpose, selective and potent agonists and antagonists for this receptor are needed. The H₃ receptor and H₄ receptor proteins are each other's closest relatives and show a relatively high level of homology (Hough, 2001), especially within the seven transmembrane domains, which are thought to bind small molecule agonists and antagonists. As might be expected on the basis of such a high homology, the H₄ receptor binds many imidazole-based H₃ receptor ligands with high affinity (Lim et al., 2005; Gbahou et al., 2006). The previously presumed H₃ receptor-selective, inverse agonist, thioperamide (Arrang et al., 1987) binds to the related H₄ receptor with equal affinity and is therefore now classified as a non-selective H₃/H₄ receptor inverse agonist (Lim et al., 2005). Some pharmaceutical companies have meanwhile started to focus on the discovery of selective non-imidazole H₄ receptor antagonists. This involvement has resulted in a number of patent applications for potent H4 receptor antagonists (Lim et al., 2006b). Currently, JNJ 7777120 (Jablonowski et al., 2003) and the related VUF 6002 (Terzioglu et al., 2004; Venable et al., 2005) should be considered as prototypic non-imidazole H₄ receptor antagonists.

Considering their high value in research, we have focused on the discovery of selective non-imidazole H₄ receptor agonists, as well. Previously, we have described the histamine analogue, 4-methylhistamine (Durant *et al.*, 1975) as a potent H₄ receptor agonist (Lim *et al.*, 2005). Moreover, we recently reported on the synthesis and initial structure–activity relationships (SAR) of the non-imidazole H₄ receptor agonist 2-(2-guanidinoethyl)isothiourea (VUF 8430) (Lim *et al.*, 2006a). In the present paper, we compare the potential of the new H₄ receptor agonist VUF 8430 as a pharmacological tool, with the standard H₄ receptor agonist 4-methylhistamine and other histamine receptor ligands. Our results clearly indicate the usefulness of VUF 8430 as a complementary pharmacological tool in dissecting the functions of the histamine H₄ receptor.

Methods

Cell culture and transfection

SK-N-MC cell lines, which stably express either the human H_3 or H_4 receptors as well as a cAMP responsive element (CRE)-driven β -galactosidase reporter gene (Lovenberg *et al.*, 1999; Liu *et al.*, 2001a), were cultured in Eagle's minimal essential

medium (EMEM) supplemented with 5% fetal calf serum, 0.1 mg·mL $^{-1}$ streptomycin, $100~\mu\text{-mL}^{-1}$ penicillin and $600~\mu\text{g·mL}^{-1}$ G418 at 37°C in 5% CO $_2$ and 95% humidity. COS-7 and HEK 293T cells were cultured in Dulbecco's modified Eagle's medium supplemented with 5% and 10%, respectively, fetal calf serum, 0.1 mg·mL $^{-1}$ streptomycin, and $100~\mu\text{-mL}^{-1}$ penicillin. Approximately 10^6 COS-7 cells were seeded in a 10 cm dish, 1 day prior to transfection. Plasmid DNA was mixed in 0.9% NaCl solution, wherafter 25 kDa polyethyleneimine (PEI) solution (1 mg·mL $^{-1}$, pH 7.0) was added to obtain a 2:1 mass ratio PEI : DNA. The mixture was incubated for 10 min, and it was then added to the COS-7 cell monolayer.

Radioligand binding assays

Cell homogenates of SK-N-MC cells expressing human H₃ receptors were incubated for 40 min at 25°C with approximately 1 nmol· L^{-1} [${}^{3}H$] N^{α} -methylhistamine in 25 mmol· L^{-1} potassium phosphate buffer and 140 mmol·L⁻¹ NaCl (pH 7.4 at room temperature), whereas cell homogenates of SK-N-MC expressing human H₄ receptors were incubated 1 h at room temperature with 10 nmol·L⁻¹ [³H]histamine in 50 mmol·L⁻¹ Tris-HCl (pH 7.4 at 37°C), with or without competing ligands. Bound radioligand was collected on 0.3% PEI-pretreated 96-well GF/C filters, which were washed three times with 3 mL of ice-cold washing buffer (4°C) containing 25 mmol·L⁻¹ Tris-HCl and 140 mmol·L⁻¹ NaCl (pH 7.4 at 4°C) for the H₃ receptor and 50 mmol·L⁻¹ Tris-HCl (pH 7.4 at 4°C) for the H₄ receptor). The binding analysis for mouse and rat H₄ receptors were performed as described above for human H₄ receptors. The binding analysis of [3H]mepyramine and [125I]iodoaminopotentidine binding to human H₁ receptors and human H₂ receptors, respectively, was performed according to Bakker et al. (2004). The binding data were analysed with Prism 4.0 (Graphpad Software, Inc.) and data are presented as mean \pm SEM.

Reporter gene assay

A CRE-β-galactosidase reporter gene assay was employed to determine the activity of the tested ligands at either the human H_3 or H_4 receptor. Approximately 4×10^6 cells per 96-well plate of SK-N-MC cells were exposed for 6 h to the tested compounds in serum-free EMEM medium containing 1 μmol·L⁻¹ forskolin. Thereafter, the medium was discarded, the cells were lysed in 100 µL assay buffer [100 mmol·L⁻¹ 4 mmol·L⁻¹ sodium phosphate buffer at pH 8.0, 2-nitrophenol-β-D-pyranoside (ONPG), 0.5% Triton X-100, $2 \text{ mmol} \cdot \text{L}^{-1} \quad \text{MgSO}_4, \quad 0.1 \text{ mmol} \cdot \text{L}^{-1} \quad \text{MnCl}_2, \quad 40 \text{ mmol} \cdot \text{L}^{-1}$ β-mercaptoethanol], incubated overnight at room temperature, and the β -galactosidase activity was determined at 420 nm with a PowerwaveX340 plate reader (Bio-Tek Instruments, Inc., USA). To measure activity of the compounds at H₂ receptors, approximately 4×10^6 resuspended HEK 293T cells were transiently cotransfected with a mixture containing 2.5 μg CRE-β-galactosidase reporter gene and 2.5 μg cDNA of the human H₂ receptor, and 35 μL of 1 mg·mL⁻¹ 25 kDa linear PEI, and transferred into a 96-well plate $(4 \times 10^4 \text{ cells per})$ well). After incubation of 24 h, the cells were exposed with the tested ligands for 6 h. The β-galactosidase activity was determined as described above. For the H₁ receptor, HEK 293T cells were contransfected with NFAT-luciferase reporter gene and cDNA of the human H₁ receptor, with a method as described above, and incubated 24 h before the cells were exposed with the tested compounds for 6 h. Subsequently, the luciferase was determined as described previously (Bakker *et al.*, 2004). The intrinsic activity of agonists was determined relative to the activity of histamine.

Generation of monocyte-derived dendritic cells

Peripheral blood mononuclear cells (PBMC) were separated from heparinized buffy coats by density gradient centrifugation on Lymphoprep (Fresenius Kabi Norge AS, Norway). Adherent cells were obtained by plastic adherence: 1×10^8 PBMC were plated in 80 cm² culture flasks (Nuclon™∆, Nunc GmbH & Co AG, Wiesbaden, Germany) in Iscove medium supplemented with 5% v/v AB serum, 1% w/v non-essential salts, 2 mmol·L⁻¹ L-glutamine, 100 μ·mL⁻¹ penicillin, 100 mg⋅mL⁻¹ streptomycin and 0.5% w/v gentamycin (all from Biochrom AG, Berlin, Germany) for 1 h (37°C, 5% CO₂, humidified atmosphere). The non-adherent cells were removed by vigorous washing with phosphate buffered saline and visual inspection. The adherent cells (enriched monocytes, purity at least 85%) were further cultured in RPMI 1640 medium supplemented with 5% v/v FCS, 12 mmol·L⁻¹ HEPES, 2 mmol·L⁻¹ L-glutamine, 100 U·mL⁻¹ penicillin, 100 mg·mL⁻¹ streptomycin, interleukin-4 (10 ng·mL⁻¹, R&D Systems, Wiesbaden, Germany) granulocyte-monocyte-colony stimulating factor (50 ng·mL⁻¹, Berlex Pharmaceutical Company, Montville, USA). Half of the medium was replaced by fresh medium on days 3 and 5 of culture. Non-adherent cells were harvested at day 7 of culture and considered as MoDC as described previously (Gutzmer et al., 2005).

Chemotaxis assay

Chemotaxis of MoDC was measured over polycarbonate membranes with 5 μm pore diameter (Corning Inc., Costar, NY, USA). CCL2 (positive control, 0.8 nmol·L $^{-1}$), histamine, 4-methylhistamine and VUF 8430 (at 10 $\mu mol\cdot L^{-1}$) were used as a chemoattractant in the lower chamber. The upper chamber with the membrane was filled with 250 μL medium containing 10^6 MoDCs. Chemotaxis was allowed for 1.5 h and the number of transmigrated cells was counted after staining with Trypan blue in a Burker-Türk counting chamber.

Gastric acid secretion in vivo

All animal procedures were in accordance with international guidelines governing animal experimentation, which was approved by the Ethics Committee of the University of Parma. Experiments were carried out with adult male Wistar rats (7–9 weeks, 200–250 g), purchased from Harlan-Italy (MI) and housed at constant temperature (20°C) and humidity (50–55%), with alternating 12-h light and dark cycles, and fed with standard laboratory chow and tap water. Gastric secretion in anaesthetized rats was measured by the stomach-lumen perfusion technique of Bertaccini *et al.* (1968) with minor modifications. The rats were used after 18-h fasting

with free access to water. After anaesthesia with ethylurethane (1.25 g·kg⁻¹ intraperitoneally), the stomach was perfused (60 mL·h⁻¹) through an oesophageal cannula with warm saline (NaCl 154 mmol·L⁻¹, 37°C, pH 5.5) at a rate of 1 mL·min⁻¹, using a peristaltic pump. The perfusion fluid was collected in 10-min periods via a duodenal cannula, and titrated to pH 7 with 10 mmol·L⁻¹ NaOH, using an automatic titration system (Radiometer, Copenhagen). In separate experiments, the histamine receptor ligands, amthamine $(0.1-1000 \, \mu \text{mol} \cdot \text{kg}^{-1})$, histamine $(0.01-1000 \, \mu \text{mol} \cdot \text{kg}^{-1})$, 4-methylhistamine (3–300 μmol·kg⁻¹), dimaprit (3–300 μmol·kg⁻¹) or VUF 8430 (30–300 μmol·kg⁻¹) were injected intravenously (i.v.) to investigate the effects on basal acid secretion. The H₂ receptor inverse agonist ranitidine (3 mg·kg⁻¹ or 8.6 μmol·kg⁻¹ i.v.) or the H₄ receptor antagonist JNJ 7777120 ($10 \text{ mg} \cdot \text{kg}^{-1}$ or $36 \,\mu\text{mol} \cdot \text{kg}^{-1}$ i.v.) was given 30 min before secretagogues. Acid responses to secretory compounds were calculated for each rat by subtracting basal acid output (average of two collection periods before the stimulant injection) from the maximal acid response (average of two collection periods) and expressed as ΔμEq HCl·kg⁻¹·min⁻¹. Amthamine, dimaprit, histamine, 4-methylhistamine, ranitidine, ethyl-urethane and VUF 8430 were dissolved in saline. DMSO 100% was used as the vehicle to dissolve JNJ 7777120. Each agent was prepared immediately before use and administered in a volume of 0.1 mL per 100 g body weight. Control animals received the vehicle in place of the active agent.

Data analysis

Results are expressed as the means \pm SEM of 5–8 rats per group. Comparisons between two groups were made by using the Student's *t*-test for unpaired data. A value of P < 0.05 was considered statistically significant. The software package Prism GraphPad 4.0 (GraphPad Software Inc., San Diego, CA) was used to process data.

Materials. Amthamine dihydrobromide, dimaprit dihydrobromide, JNJ 7777120, 4-methylhistamine dihydrochloride, thioperamide fumarate and VUF 8430 [S-(2-guanidylethyl)isothiourea dihydrobromide] were synthesized at the Department of Medicinal Chemistry, Vrije Universiteit Amsterdam, while [125] iodoaminopotentidine was labelled at the Department of Nuclear Medicine and PET Research, Vrije Universiteit Medical Centre, Amsterdam. Forskolin, histamine dihydrochloride, mepyramine (pyrilamine maleate), N^{α} -methylhistamine dihydrochloride, Pertussis 750 kDa PEI, ranitidine hydrochloride, agmatine sulfate, putrescine dihydrochloride, spermine, spermidine and Trypan blue were purchased from Sigma RBI (USA). ONPG and G418 were purchased from Duchefa (The Netherlands), and [³H]N^α-methylhistamine (85 Ci⋅mmol⁻¹), [³H]histamine (12.4 Ci·mmol⁻¹), [³H]mepyramine (23 Ci·mmol⁻¹) were from Perkin-Elmer Life Science, Inc. (USA), and 25 kDa PEI (for transfection) was from Polyscience, Inc. (Germany). Gifts of SK-N-MC cell lines expressing human H₃ receptors, human H₄ receptors, mouse H4 receptors, rat H3 receptors and rat H4 receptors from Dr Lovenberg are greatly acknowledged (Liu et al., 2001a,b).

Results

Binding of VUF 8430 and related compounds to human and rodent H₄ receptors

In radioligand binding studies using [3 H]histamine and SK-N-MC cells stably expressing the human H $_{4}$ receptor (Liu et al., 2001a), the newly discovered VUF 8430 (Lim et al., 2006a) was almost as potent as histamine and the known histamine H $_{4}$ receptor agonist, 4-methylhistamine (Lim et al., 2005) in binding to the H $_{4}$ receptor (Figure 1A, Table 1). Moreover, the structurally related H $_{2}$ receptor agonist dimaprit was approximately 100-fold less effective than VUF 8430 in displacing [3 H]histamine binding to the human H $_{4}$ receptor (Figure 1A, Table 1).

The endogenous amine, agmatine, has a clear structural resemblance to VUF 8430 (Table 1) and is formed by decarboxylation of L-arginine by arginine decarboxylase (ADC) and hydrolyzed by agmatinase to putrescine. Agmatine binds to several receptors in the brain and has been proposed as a novel neurotransmitter (Li *et al.*, 1994; Reis and Regunathan, 2000). In our experiments, agmatine showed a micromolar affinity for the human H_4 receptor (Figure 1A, Table 1). As can be seen in Figure 1A, its amino acid precursor, arginine, was totally inactive at the H_4 receptor, whereas its metabolite, putrescine, still showed moderate affinity (Figure 1B, Table 1). Following this observation, we tested a variety of other polyamines (Figure 1B) and found that the putrescine homologue cadaverine, spermine and spermidine all showed

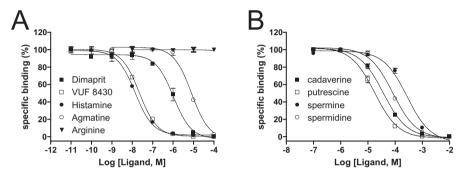


Figure 1 Binding of a variety of amines to the human H_4 receptor. (A) Histamine, dimaprit, VUF 8430 and agmatine, but not arginine, and (B) the polyamines putrescine, cadaverine, spermidine and spermine dose-dependently displaced binding of [3H]histamine to the human H_4 receptor stably expressed in SK-N-MC cells.

Table 1 Affinity (pK₁) of H₄ receptor ligands for different species variants of H₄ receptor stably expressed in SK-N-MC cells

Ligand	Structure	Human	Mouse	Rat
Histamine	NH ₂	7.9 ± 0.1	7.4 ± 0.1	7.3 ± 0.1
4-methyl-histamine	HN NH ₂	7.6 ± 0.1	7.2 ± 0.1	6.7 ± 0.1
Dimaprit	HN NH	6.8 ± 0.1	6.2 ± 0.1	6.0 ± 0.1
VUF 8430	H ₂ N H NH ₂	7.5 ± 0.1	7.0 ± 0.1	6.9 ± 0.1
Agmatine	H_2N N N N N N N N N	5.6 ± 0.1	5.1 ± 0.1	5.1 ± 0.2
L-arginine	NH COOH H ₂ N NH ₂	<3	n.d.	n.d.
Putrescine	II NH H ₂ N NH ₂	4.9 ± 0.1	n.d.	n.d.
Cadaverine	H_2N NH_2	4.7 ± 0.1	n.d.	n.d.
Spermidine	NH ₂ -(CH ₂) ₄ -NH-(CH ₂) ₃ -NH ₂	3.7 ± 0.1	n.d.	n.d.
Spermine	NH ₂ -(CH ₂) ₃ -NH-(CH ₂) ₄ -NH-(CH ₂) ₃ -NH ₂	4.3 ± 0.1	n.d.	n.d.

Data shown are mean \pm SEM of at least three independent experiments. n.d., not determined.

moderate to weak affinities for the human H_4 receptor (Figure 1B, Table 1).

Agonist activities of VUF 8430 and agmatine at human histamine receptors

We subsequently evaluated the functional activities of histamine, VUF 8430, 4-methylhistamine and agmatine at the four human histamine receptor subtypes. As reported previously (Lim *et al.*, 2005; 2006a), both histamine ($\alpha = 1$ by definition), 4-methylhistamine ($\alpha = 1$) and VUF 8430 ($\alpha = 1$) act as full agonists at the human H₄ receptor as measured by the G_i-protein mediated inhibition of the forskolin-induced (1 μmol·L⁻¹) CRE-mediated transcription of β-galactosidase in SK-N-MC cells, stably expressing the human H₄ receptor $(B_{max} = 1.8 \text{ pmol} \cdot \text{mg}^{-1} \text{ protein})$ and a CRE- β -galactosidase reporter gene (Figure 2A). In this assay, the endogenous agonist histamine was slightly more potent (pEC₅₀ = 7.7 \pm 0.1, n = 16) compared with VUF 8430 (pEC₅₀ = 7.3 ± 0.1, n=6) or 4-methylhistamine (pEC₅₀ = 7.4 \pm 0.1, n=5). Agmatine, under these experimental conditions, was a partial agonist ($\alpha = 0.65 \pm 0.05$, n = 4) with a pEC₅₀ value of 5.4 ± 0.1 (n = 4). At the related human H₃ receptor (Figure 2B), both histamine ($\alpha = 1$ by definition) and VUF 8430 ($\alpha = 1$) were full agonists, as measured by the inhibition of the 1 μmol·L⁻¹ forskolin-induced CRE-mediated transcription of β-galactosidase in SK-N-MC cells stably expressing the human H₃ receptor (B_{max} = 475 fmol⋅mg⁻¹ protein) and a CREβ-galactosidase reporter gene. Agmatine also acts as a full agonist ($\alpha = 1$) at the H₃ receptor (Figure 2B) with a pEC₅₀ value of 6.1 \pm 0.1 (n = 3); 4-methylhistamine was clearly less effective at the human H₃ receptor (Figure 2B) and was a partial agonist ($\alpha = 0.38 \pm 0.12$, n = 3) with a pEC₅₀ value of 6.1 ± 0.1 (n = 3).

VUF 8430 did not interact with the histamine H_1 and H_2 receptor subtypes. Histamine displayed a pEC₅₀ value of 7.0 ± 0.1 (n = 4) at human H_1 receptors, measured using HEK 293T cells transiently expressing the human H_1 receptor and a NFAT-luciferase reporter gene (Figure 2C). In the same experimental set-up, 4-methylhistamine, VUF 8430 and agmatine showed no agonist activity at the H_1 receptor at concentrations up to 100 μmol·L⁻¹ (Figure 2C). At human H_2 receptors, histamine displayed a pEC₅₀ of 7.2 ± 0.1 (α = 1, n = 4), measured using COS-7 cells transiently expressing the human H_2 receptor and a CRE-β-galactosidase reporter gene. In this assay, 4-methylhistamine (α = 1) exhibited a pEC₅₀ of 6.8 ± 0.1 (n = 4), whereas VUF 8430 and agmatine only weakly activated the human H_2 receptor at 100 μmol·L⁻¹ (Figure 2D).

Affinity of H_4 receptor agonists at rodent histamine receptors Previously, significant species differences were reported for the binding of histamine to rodent H_4 receptors (Liu *et al.*, 2001b). We therefore tested VUF 8430 and agmatine for their affinity at mouse and rat H_4 receptors (Figure 3A,B, Table 1). As previously reported (Liu *et al.*, 2001b), histamine showed an three- to fivefold lower affinity for the mouse and rat H_4 receptor compared with the human H_4 receptor, measured by displacement of [3H]histamine binding to homogenates of SK-N-MC cells stably expressing the various H_4 receptor orthologs (Figure 3A, Table 1). Similarly, both VUF 8430 and

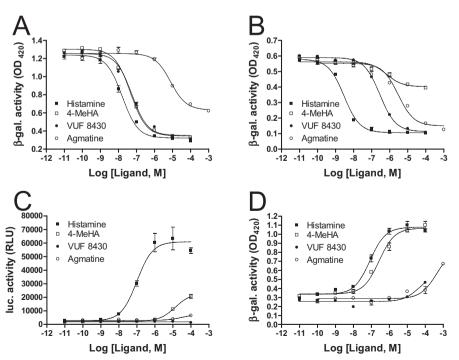


Figure 2 Functional activity of histamine, 4-methylhistamine (4-MeHA), VUF 8430 and agmatine at the human H_4 receptor (A), human H_3 receptor (B) expressed in SK-N-MC cells and human H_1 receptor (C) and human H_2 receptor (D) expressed in HEK 293T cells. The NFAT-luciferase reporter gene assay was used to measure H_1 receptor activity, while the CRE-β-galactosidase was employed to measure activation of cAMP production by H_2 receptor or inhibition of forskolin-induced cAMP generation by the H_3 and H_4 receptors.

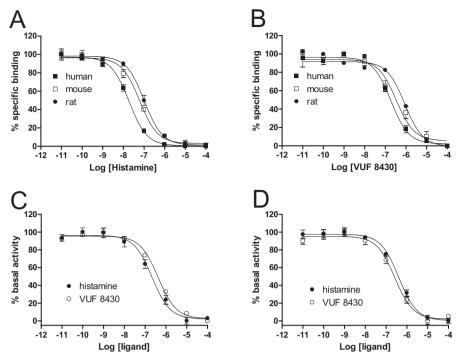


Figure 3 Interaction of VUF 8430 with the H_4 receptor species variants. Histamine (A) and VUF 8430 (B) inhibits binding of [3H]histamine to human, mouse and rat H_4 receptors stably expressed in SK-N-MC cells. VUF 8430 inhibited forskolin-induced cAMP responsive element (CRE) through mouse H_4 receptor (C) and rat H_4 receptor (D), to the same extent as histamine.

Table 2 Affinity (pK_i) of H₄ receptor ligands at rat histamine receptors

Ligand	pK _i at rat histamine receptor				
	H ₁	H ₂	Н₃	H ₄	
Histamine	4.5 ± 0.2	4.3 ± 0.1	8.1 ± 0.1	7.4 ± 0.1	
4-methylhistamine	<4	4.1 ± 0.1	5.2 ± 0.1	6.8 ± 0.1	
Dimaprit	4.4 ± 0.2	4.5 ± 0.3	6.3 ± 0.1	6.1 ± 0.1	
VUF 8430	<4	3.7 ± 0.1	6.5 ± 0.1	6.9 ± 0.1	
Thioperamide	4.1 ± 0.1	3.7 ± 0.1	7.8 ± 0.1	7.2 ± 0.1	
INI 7777120	5.1 ± 0.1	4.9 ± 0.1	5.3 ± 0.1	7.8 ± 0.1	
VUF 6002	4.7 ± 0.1	4.5 ± 0.1	5.3 ± 0.1	6.8 ± 0.1	

Data shown are mean \pm SEM of at least three independent experiments.

agmatine showed a somewhat lower affinity at the rodent receptor (Figure 3B, Table 1). At both the mouse (Figure 3C) and rat H₄ receptor (Figure 3D), VUF 8430 was as effective as histamine in inhibiting the 1 μ mol·L⁻¹ forskolin-induced CRE activation, displaying pEC₅₀ values of 6.5 \pm 0.1 (α = 1, n = 3) and 6.4 \pm 0.1 (α = 1, n = 4) respectively. Furthermore, agmatine equipotently and partially activates the mouse and rat H₄ receptors with a pEC50 value of 4.6 \pm 0.1.

To understand the action of the various $\rm H_4$ receptor agonists in *in vivo* rodent models we have also tested histamine, VUF 8430 and 4-methylhistamine at the four different rat histamine receptors (Table 2). As expected, both 4-methylhistamine and VUF 8430 do not show significant affinity (pK_i < 4) for the rat $\rm H_1$ receptor and each display a similar affinity for the rat $\rm H_2$ receptor compared with histamine (Table 2). As observed for the human $\rm H_3$ receptor, VUF 8430 binds with higher affinity to rat $\rm H_3$ receptors, compared with

4-methylhistamine, whereas both H_4 receptor agonists show a similar affinity for rat H_4 receptors (Table 2). In the same set of experiments, we also tested the H_4 receptor antagonists thioperamide, JNJ 7777120 (Jablonowski *et al.*, 2003) and VUF 6002 (Terzioglu *et al.*, 2004; Venable *et al.*, 2005) at the four rat histamine receptors. As observed previously for the human histamine receptors (Lim *et al.*, 2005), thioperamide was slightly more active at the rat H_3 receptor (pK_i = 7.8) compared with the rat H_4 receptor (pK_i = 7.2), whereas JNJ 7777120 and VUF 6002 showed a good selectivity (>100 fold) for the rat H_4 receptor (Table 2).

In vivo evaluation of VUF 8430 as a pharmacological tool for studies of H_4 receptors

In view of the difference in agonist activity at H_2 receptors, of 4-methylhistamine and VUF 8430 (Figure 2D) we compared

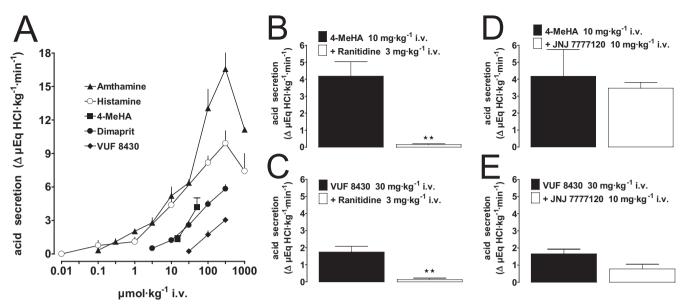


Figure 4 Analysis of gastric acid secretion in anaesthetized rats. The histamine H₂ receptor agonists amthamine and 4-methylhistamine (4-MeHA) induced acid secretion, but VUF 8430 only showed minimal effects at high doses (A). The effects of 4-methylhistamine (30 mg·kg⁻¹ or 93 μmole·kg⁻¹ i.v.) (B) or VUF 8430 (30 mg·kg⁻¹ or 50 μmole·kg⁻¹ i.v.) (C) were inhibited by ranitidine, but no significant inhibition by JNJ 7777120 (D and E) was observed. Values represent the mean \pm SEM of responses in 6–8 animals for each experimental group. Comparisons between two groups were made by using the Student's *t*-test for unpaired data. ***P* < 0.01 versus 4-methylhistamine (B) or VUF 8430 (C).

VUF 8430 and 4-methylhistamine for their potential to stimulate H₂ receptor function in vivo. We therefore measured gastric acid secretion in unconscious rats after application of histamine, the H₂ receptor agonists dimaprit and amthamine, VUF 8430, or 4-methylhistamine. Both histamine and amthamine are potent inducers in vivo of gastric acid secretion in the rat (Figure 4A). Moreover, dimaprit and 4methylhistamine also induce gastric acid secretion, although higher doses were needed, whereas VUF 8430 only marginally induced gastric acid secretion at the highest tested doses (Figure 4A). These data are in good accordance with our in vitro findings in transfected cells (Figure 2D). Gastric acid secretion induced by VUF 8430 (30 mg·kg⁻¹ or 50 µmole·kg⁻¹, i.v.) and 4-methylhistamine (10 mg·kg⁻¹ or 93 μmole·kg⁻¹, i.v.) was suppressed by administration of the selective H₂ receptor antagonist ranitidine (3 mg·kg⁻¹, i.v.) (Figure 4B). The H₄ receptor antagonist JNJ 7777120 was not able to reduce 4-methylhistamine or VUF 8430-induced gastric acid secretion (Figure 4B).

Ex vivo evaluation of VUF 8430

We also evaluated the $\rm H_4$ receptor-mediated effects of VUF 8430 in an *ex vivo* assay of migration of human MoDCs. The $\rm H_4$ receptor has been shown to be expressed on the MoDCs and plays a role in histamine-induced chemotaxis (Gutzmer *et al.*, 2005). Histamine-induced migration of MoDCs as efficaciously as the chemokine, CCL2 and this effect was also mimicked by $\rm H_4$ receptor agonists 4-methylhistamine and VUF 8430 (Figure 5).

Discussion and conclusions

In this study, we evaluated VUF 8430 as a pharmacological tool for the study of histamine H_4 receptors, using *in vitro*,

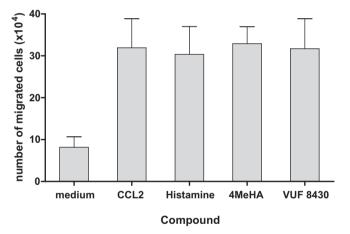


Figure 5 Chemotaxis of human monocyte-derived dendritic cells (MoDCs). MoDCs were loaded in upper wells of migration chambers and allowed to migrate towards chemotactic agents in the lower well through polycarbonate membrane with 5- μ m pore diameter. The positive control (0.8 nmol·L⁻¹ CCL2) and the tested compounds [histamine, 4-methylhistamine (4-MeHA), and VUF 8430, all at 10 μ mol·L⁻¹] were diluted in cell culture medium as described in *Methods*.

ex vivo and in vivo models. VUF 8430 was originally synthesized in a receptor programme looking for close analogues of the $\rm H_2$ receptor agonist dimaprit (Durant et al., 1977). However, with the discovery of new histamine receptor subtypes ($\rm H_3$ and $\rm H_4$ receptors), we have recently tested a large number of histamine receptor ligands for their activity at the human $\rm H_4$ receptor (Lim et al., 2005). This detailed evaluation resulted in the discovery of the moderately potent $\rm H_2$ receptor agonist, 4-methylhistamine, as a potent $\rm H_4$ receptor agonist (Lim et al., 2005). Moreover, we also observed that dimaprit

showed reasonable H₄ receptor agonist activity (Lim *et al.*, 2005) and subsequently described a new efficient synthetic pathway and an initial SAR of the related VUF 8430 (Lim *et al.*, 2006a). This dimaprit derivative shows high affinity at the human H₄ receptor and acts as a full agonist (Lim *et al.*, 2006a; this study). As shown in this study, both 4-methylhistamine and VUF 8430 also act as H₄ receptor agonists at endogenously expressed H₄ receptors in MoDCs. Expression of H₄ receptors was detected in MoDCs at both the mRNA and protein level and these receptors were involved in the activation of the transcription factor AP-1, the inhibition of IL-12p70 secretion and chemotaxis (Gutzmer *et al.*, 2005). We found that both VUF 8430 and 4-methylhistamine induced chemotaxis of MoDCs as effectively as histamine or the chemokine CCL2.

The discovery of VUF 8430 as an H₄ receptor agonist led us to investigate whether agmatine is an H4 receptor ligand as well. Agmatine is a metabolite of L-arginine, occurs naturally in the body and has been considered to act as an important chemical messenger at other sites such as imidazoline receptors and α₂-adrenoceptors (Li et al., 1994; Reis and Regunathan, 2000; Raasch et al., 2001). In our study, we showed that replacement of the isothiourea group of VUF 8430 with an amine function, as in agmatine, resulted in a sharp drop in binding affinity for H₄ receptors. Nevertheless, in contrast to its precursor, L-arginine, agmatine binds to the H₄ receptor with a K_i value of 2 μmol·L⁻¹. In the reporter gene assay, agmatine showed partial agonist activity at the human H₄ receptor and full agonism at human H₃ receptors. Micromolar concentrations of agmatine are considered to have physiological importance (Lortie et al., 1996; Raasch et al., 2001) and are found in the brain, kidney and other tissues (Li et al., 1994; Lortie et al., 1996; 2004). Hence, agmatine may be one of the endogenous ligands for the H₃ and H₄ receptor. These data also show the importance of the isothiourea group of VUF 8430 in the binding to H₄ receptors. Previously, we reported that the isothiourea group most likely forms hydrogen bonds with residues E5.46 (1.82 Å) and S6.52 (2.21 Å) in the receptor protein (Jongejan et al., 2008). The resulting H-bonding network is identical to the pattern observed for the imidazole ring of histamine, and an essential structural feature for the binding of histamine to the H₄ receptor (Lim et al., 2005).

Besides agmatine, we also observed that other polyamines bind to H₄ receptors; putrescine, cadaverine, spermidine and spermine all showed affinity for H₄ receptors. Cadaverine is commonly found in microorganisms, while putrescine, spermidine and spermine are metabolites of arginine endogenously found in the human body. Like agmatine, concentrations of the polyamines are tightly controlled and they increase under certain conditions, such as liver regeneration, sepsis, brain ischaemia and acute excitotoxic brain damage (Anehus *et al.*, 1986; Gilad *et al.*, 1996; Noguchi *et al.*, 1996; Vivo *et al.*, 2002). The relevance of this action of polyamines on H₄ receptor remains to be elucidated.

A detailed *in vitro* comparison of the actions of VUF 8430, agmatine and the earlier identified H_4 receptor agonist 4-methylhistamine, indicate that all three compounds, like histamine (Liu *et al.*, 2001b), show a somewhat reduced affinity at the rat and mouse H_4 receptors compared with the

human H₄ receptor. In radioligand binding and functional studies, VUF 8430 also shows reasonable affinity and full agonist efficacy at the related H₃ receptors. Similarly, agmatine acts as an efficacious agonist at the H₃ receptor, whereas 4-methylhistamine only shows some agonistic action at H₃ receptors, at high concentrations. Both VUF 8430 and agmatine are very weakly active at the H₂ receptor and inactive at the H₁ receptor. In contrast, despite the relatively low affinity of 4-methylhistamine at the H₂ receptor, 4-methylhistamine is as active as histamine at this receptor. Interestingly, the differential effects of 4-methylhistamine and VUF 8430 at the H₂ receptor were also observed in vivo. In anaesthetized rats, we observed that 4-methylhistamine was effective in stimulating gastric acid secretion in vivo, thus confirming a considerable activity at the H₂ receptor (Durant et al., 1975). Compared with 4-methylhistamine, VUF 8430 is considerably less potent and less efficacious in triggering gastric acid secretion in the rat. Only at very high dosages, VUF 8430 stimulated gastric acid secretion and then only to a very small extent. The stimulatory effect of both H₄ receptor agonists was effectively blocked by the H₂ receptor inverse agonist ranitidine, but not by the H₄ receptor antagonist JNJ 7777120. Based on these data, we conclude that 4-methylhistamine in vivo shows considerable H2 receptor agonistic activities at dosages >3 mg·kg⁻¹ i.v. For VUF 8430 only at considerable higher dosages (30 mg·kg⁻¹ i.v.) some minor H₂ receptormediated effects were observed in vivo.

In conclusion, our search for new ligands for the histamine H₄ receptor has resulted in the discovery of the potent H₄ receptor agonist VUF 8430, which shows a pharmacological profile clearly different from that of the other high affinity receptor agonist, 4-methylhistamine. 4-methylhistamine still showed considerable H₂ receptor agonistic activity (in vitro and in vivo), but was devoid of H₁ receptor activity and only weakly activated the H₃ receptor, VUF 8430 on the other hand showed a limited selectivity towards the H₃ receptor, but was very selective with respect to the H₁ or H₂ receptors. Consequently, both high affinity H₄ receptor agonists are complementary and together with the available selective H4 receptor antagonists can serve as pharmacological tools in future studies to validate the H₄ receptor as a new drug target. Furthermore, identification of VUF 8430 also led to the discovery of agmatine as potential endogenous H₄ receptor agonist. The physiological relevance of agmatine and the polyamines as histamine H₄ receptor ligands remains to be elucidated.

Acknowledgements

We thank the Technology Foundation (Stichting voor de Technische Wetenschapppen) of the Netherlands Foundation of Scientific Research (Nederlandse Organisatie voor Wetenschappelijk Onderzoek) for financial support through a PIONIER award to Rob Leurs.

Conflict of interest

None.

References

- Alexander SPH, Mathie A, Peters JA (2008). Guide to receptors and channels (GRAC), 3rd edn. *Br J Pharmacol* 153 (Suppl. 2): S1–S209.
- Anehus S, Yngner T, Hafstrom L, Heby O (1986). Increased urinary polyamine excretion during liver regeneration. *Biochem Med Metab Biol* 35: 322–326.
- Arrang JM, Garbarg M, Lancelot JC, Lecomte JM, Pollard H, Robba M *et al.* (1987). Highly potent and selective ligands for histamine H3-receptors. *Nature* **327**: 117–123.
- Bakker RA, Weiner DM, ter Laak T, Beuming T, Zuiderveld OP, Edelbroek M *et al.* (2004). 8R-lisuride is a potent stereospecific histamine H1-receptor partial agonist. *Mol Pharmacol* **65**: 538–549.
- Bertaccini G, Endean R, Erspamer V, Impicciatore M (1968). The actions of caerulein on gastric acid secretion of the dog and the rat. *Br J Pharmacol* **34**: 311–329.
- Cianchi F, Cortesini C, Schiavone N, Perna F, Magnelli L, Fanti E *et al.* (2005). The role of cyclooxygenase-2 in mediating the effects of histamine on cell proliferation and vascular endothelial growth factor production in colorectal cancer. *Clin Cancer Res* 11: 6807–6815.
- Dunford PJ, O'Donnell N, Riley JP, Williams KN, Karlsson L, Thurmond RL (2006). The histamine H4 receptor mediates allergic airway inflammation by regulating the activation of CD4+ T cells. *J Immunol* 176: 7062–7070.
- Dunford PJ, Williams KN, Desai PJ, Karlsson L, McQueen D, Thurmond RL (2007). Histamine H(4) receptor antagonists are superior to traditional antihistamines in the attenuation of experimental pruritus. *J Allergy Clin Immunol* 119: 176–183.
- Durant GJ, Ganellin CR, Parsons ME (1975). Chemical differentiation of histamine H1- and H2-receptor agonists. *J Med Chem* **18**: 905–909.
- Durant GJ, Ganellin CR, Parsons ME (1977). Dimaprit, (S-[3-(N,N-dimethylamino)propyl]isothiourea). A highly specific histamine H2-receptor agonist. Part 2. Structure-activity considerations. *Agents Actions* 7: 39–43.
- de Esch IJ, Thurmond RL, Jongejan A, Leurs R (2005). The histamine H4 receptor as a new therapeutic target for inflammation. *Trends Pharmacol Sci* **26**: 462–469.
- Gantner F, Sakai K, Tusche MW, Cruikshank WW, Center DM, Bacon KB (2002). Histamine h(4) and h(2) receptors control histamine-induced interleukin-16 release from human CD8(+) T cells. *J Pharmacol Exp Ther* 303: 300–307.
- Gbahou F, Vincent L, Humbert-Claude M, Tardivel-Lacombe J, Chabret C, Arrang JM (2006). Compared pharmacology of human histamine H(3) and H(4) receptors: structure–activity relationships of histamine derivatives. *Br J Pharmacol* **147**: 744–754.
- Gilad GM, Gilad VH, Rabey JM (1996). Arginine and ornithine decarboxylation in rodent brain: coincidental changes during development and after ischemia. *Neurosci Lett* **216**: 33–36.
- Gutzmer R, Diestel C, Mommert S, Kother B, Stark H, Wittmann M *et al.* (2005). Histamine H4 receptor stimulation suppresses IL-12p70 production and mediates chemotaxis in human monocyte-derived dendritic cells. *J Immunol* 174: 5224–5232.
- Hofstra CL, Desai PJ, Thurmond RL, Fung-Leung WP (2003). Histamine H4 receptor mediates chemotaxis and calcium mobilization of mast cells. *J Pharmacol Exp Ther* 305: 1212–1221.
- Hough LB (2001). Genomics meets histamine receptors: new subtypes, new receptors. *Mol Pharmacol* **59**: 415–419.
- Jablonowski JA, Grice CA, Chai W, Dvorak CA, Venable JD, Kwok AK et al. (2003). The first potent and selective non-imidazole human histamine H4 receptor antagonists. J Med Chem 46: 3957–3960.
- Jongejan A, Lim HD, Smits RA, de Esch IJ, Haaksma E, Leurs R (2008). Delineation of agonist binding to the human histamine h4 receptor using mutational analysis, homology modeling, and ab initio calculations. J Chem Inf Model 48: 1455–1463.

- Leurs R, Bakker RA, Timmerman H, de Esch IJ (2005). The histamine H3 receptor: from gene cloning to H3 receptor drugs. *Nat Rev Drug Discov* 4: 107–120.
- Li G, Regunathan S, Barrow CJ, Eshraghi J, Cooper R, Reis DJ (1994). Agmatine: an endogenous clonidine-displacing substance in the brain. *Science* **263**: 966–969.
- Lim HD, van Rijn RM, Ling P, Bakker RA, Thurmond RL, Leurs R (2005). Evaluation of histamine H1-, H2-, and H3-receptor ligands at the human histamine H4 receptor: identification of 4-methylhistamine as the first potent and selective H4 receptor agonist. *J Pharmacol Exp Ther* 314: 1310–1321.
- Lim HD, Smits RA, Bakker RA, van Dam CM, de Esch IJ, Leurs R (2006a). Discovery of S-(2-guanidylethyl)-isothiourea (VUF 8430) as a potent nonimidazole histamine H4 receptor agonist. *J Med Chem* 49: 6650–6651.
- Lim HD, Smits RA, Leurs R, De Esch IJ (2006b). The emerging role of the histamine H4 receptor in anti-inflammatory therapy. *Curr Top Med Chem* 6: 1365–1373.
- Ling P, Ngo K, Nguyen S, Thurmond RL, Edwards JP, Karlsson L *et al.* (2004). Histamine H4 receptor mediates eosinophil chemotaxis with cell shape change and adhesion molecule upregulation. *Br J Pharmacol* 142: 161–171.
- Liu C, Ma X, Jiang X, Wilson SJ, Hofstra CL, Blevitt J et al. (2001a). Cloning and pharmacological characterization of a fourth histamine receptor (H(4)) expressed in bone marrow. Mol Pharmacol 59: 420–426
- Liu C, Wilson SJ, Kuei C, Lovenberg TW (2001b). Comparison of human, mouse, rat, and guinea pig histamine H4 receptors reveals substantial pharmacological species variation. *J Pharmacol Exp Ther* 299: 121–130.
- Lortie MJ, Novotny WF, Peterson OW, Vallon V, Malvey K, Mendonca M *et al.* (1996). Agmatine, a bioactive metabolite of arginine. Production, degradation, and functional effects in the kidney of the rat. *J Clin Invest* 97: 413–420.
- Lortie MJ, Satriano J, Gabbai FB, Thareau S, Khang S, Deng A et al. (2004). Production of arginine by the kidney is impaired in a model of sepsis: early events following LPS. Am J Physiol Regul Integr Comp Physiol 287: R1434–R1440.
- Lovenberg TW, Roland BL, Wilson SJ, Jiang X, Pyati J, Huvar A *et al.* (1999). Cloning and functional expression of the human histamine H3 receptor. *Mol Pharmacol* 55: 1101–1107.
- Morse KL, Behan J, Laz TM, West RE Jr, Greenfeder SA, Anthes JC *et al.* (2001). Cloning and characterization of a novel human histamine receptor. *J Pharmacol Exp Ther* **296**: 1058–1066.
- Noguchi Y, Meyer T, Tiao G, Fischer JE, Hasselgren PO (1996). Sepsis increases putrescine concentration and protein synthesis in mucosa of small intestine in rats. *Shock* 5: 333–340.
- Oda T, Morikawa N, Saito Y, Masuho Y, Matsumoto S (2000). Molecular cloning and characterization of a novel type of histamine receptor preferentially expressed in leukocytes. *J Biol Chem* **275**: 36781–36786.
- Ohki E, Suzuki M, Aoe T, Ikawa Y, Negishi E, Ueno K (2007). Expression of histamine H4 receptor in synovial cells from rheumatoid arthritic patients. *Biol Pharm Bull* 30: 2217–2220.
- Parsons ME, Ganellin CR (2006). Histamine and its receptors. *Br J Pharmacol* **147** (Suppl. 1): S127–S135.
- Raasch W, Schafer U, Chun J, Dominiak P (2001). Biological significance of agmatine, an endogenous ligand at imidazoline binding sites. *Br J Pharmacol* **133**: 755–780.
- Reis DJ, Regunathan S (2000). Is agmatine a novel neurotransmitter in brain? *Trends Pharmacol Sci* 21: 187–193.
- Takeshita K, Sakai K, Bacon KB, Gantner F (2003). Critical role of histamine H4 receptor in leukotriene B4 production and mast cell-dependent neutrophil recruitment induced by zymosan in vivo. *J Pharmacol Exp Ther* **307**: 1072–1078.
- Terzioglu N, van Rijn RM, Bakker RA, De Esch IJ, Leurs R (2004). Synthesis and structure–activity relationships of indole and benz-

- imidazole piperazines as histamine H(4) receptor antagonists. *Bioorg Med Chem Lett* 14: 5251–5256.
- Thurmond RL, Desai PJ, Dunford PJ, Fung-Leung WP, Hofstra CL, Jiang W *et al.* (2004). A potent and selective histamine H4 receptor antagonist with anti-inflammatory properties. *J Pharmacol Exp Ther* **309**: 404–413.
- Venable JD, Cai H, Chai W, Dvorak CA, Grice CA, Jablonowski JA *et al.* (2005). Preparation and biological evaluation of indole, benzimidazole, and thienopyrrole piperazine carboxamides: potent human histamine h(4) antagonists. *J Med Chem* **48**: 8289–8298.
- Vivo M, Camon L, de Vera N, Martinez E (2002). Extracellular putrescine content after acute excitotoxic brain damage in the rat. *Neurosci Lett* **330**: 74–78.
- Zhu Y, Michalovich D, Wu H, Tan KB, Dytko GM, Mannan IJ *et al.* (2001). Cloning, expression, and pharmacological characterization of a novel human histamine receptor. *Mol Pharmacol* **59**: 434–441